

OPINION

To What Extent Does Ozone Therapy Need a Real Biochemical Control System? Assessment and Importance of Oxidative Stress

Frank Antonio Hernández

Ozone Research Center Habana, Ciudad de la Habana, Cuba

Received for publication November 7, 2006; accepted March 20, 2007 (ARCMED-D-06-00475).

Ozone therapy is not officially allowed in many countries, but private medical services are using this therapy worldwide. However, appropriate control systems to assess the benefits and risks of systemic ozone therapy are not always used and in such cases the treatment is based on anecdotal reports. Oxidative stress phenomenon is becoming a highlighted biological process for ozone therapy because it is deeply involved in its mechanism of action. On the contrary, ozone therapy is an efficient regulator of the oxidative stress processes. In terms of therapeutic effects, it is convenient to know the metabolic status of the organism to face new oxidative challenges before and during ozone therapy applications. Oxidative stress is also important because it is involved as a cause or effect of many diseases. Since the 1990s, there has been the necessity of developing reliable systems for measuring oxidative stress in humans. In this sense, we have proposed a system for oxidative stress diagnosis that can serve as a control system for systemic ozone therapy applications. The system is based on the blood measurement of eight biomarkers (GSH, GPx, GST, SOD, CAT, DC, SRATB, and HPT) and the interpretation of these values by a computer-developed algorithm yielding four new indices (total antioxidant activity, total prooxidant activity, redox index and grade of oxidative stress). The system shows the patient's redox status and estimation of the oxidative stress level, with this information being relevant regarding implications on dosage and therapeutic effectiveness of ozone therapy. © 2007 IMSS. Published by Elsevier Inc.

Key Words: Ozone therapy, Oxidant state, Antioxidant state, Antioxidant enzymes, Oxidative stress.

Introduction

One of the reasons why ozone in medicine has not been approved as a common practice is its use without an appropriate control. Therefore, a safety and therapeutic control system becomes a matter of great relevance for applications in systemic ozone therapy. The use of specific biochemical parameters to control the applicable dose in terms of safety and therapeutic effects has been gaining importance (1,2) and would support the bases for nonempiric application. However, there is not yet a generalized criterion of those

specific biochemical indices that may be useful in assessing the safety and therapeutic effect of systemic ozone therapy.

Ozone Therapy and Oxidative Stress

The main ozone therapy mechanism of action is based on an extremely transitory and regulated oxidative stress imposed *ex vivo* (3). At the same time, ozone therapy acts as an efficient oxidative stress regulator stimulating the antioxidant system of the cell. Therefore, the oxidative stress phenomenon is deeply involved in ozone therapy and a test or system for assessing this process will be of great relevance. As reactive oxygen species (ROS) attack a variety of organic substrates, oxidative stress can be evaluated by measuring reaction products of oxidative damage. Hence, it is convenient before and during application of ozone therapy to know the patient's redox status in order to control the safety doses

Address reprint requests to: Frank Antonio Hernández, Dr. Sc., Departamento de Biomedicina, Centro de Investigaciones del Ozono, Centro Nacional de Investigaciones Científicas, Apdo. 6412, Ciudad Habana, Cuba; E-mail: hernandez@cnic.edu.cu

of ozone to be applied in each application. Also, in terms of therapeutic effects, it is convenient to know the metabolic status of the organism to face new oxidative challenges before and during ozone therapy applications.

The comprehensive term “oxidative stress” is currently used to indicate a complex series of biochemical processes that can affect living matter under several distinct conditions and produce both physiological and pathological effects. Oxidative stress can be defined as the prevalence within the living cell of oxidizing species or activities over the cellular antioxidant defenses, and it is this pathological perspective that has received great attention over the past decades (4,5). Until recently, a considerable amount of research has, in fact, focused on the role of oxidative stress in mediating the cell-damaging effects of a wide range of prooxidant agents (chemicals, drugs, pollutants). Several detailed reviews dealing with the mechanistic aspects of these cytotoxic processes have been published (6–9).

On the other hand, a number of studies in recent years have highlighted the role of oxidation/reduction (redox) reactions in the regulation of molecular mechanisms involved in cellular signal transduction (10–14). It is becoming clear that low physiological levels of oxidant agents (transitory or regulated oxidative stress) can have physiological roles within cells (15). This novel perspective is particularly intriguing, as it will surely advance our understanding of the role played by redox processes in human disease. Oxidative stress, if often partly or totally, is a cause of various diseases such as cancer, atherosclerosis, cataract, maculopathy, Alzheimer, Parkinson and others, but a great number diseases such as diabetes, chronic renal insufficiency, and infectious processes such as AIDS or viral hepatitis create a secondary ROS overproduction that worsens the evolution of the diseases (16,17).

Monitoring oxidative stress status becomes an interesting challenge for clinical biochemists. In the year 1990, 16 scientists from the U.S. and Canada gathered in Rockville, MD to participate in a symposium for exchanging ideas on potential methods of measuring the oxidative stress status of humans. This workshop did not reach a consensus, perhaps because it was the first effort to organize thinking about this field (18). However, it points out the necessity of developing reliable measures of oxidative stress in humans; in this sense, a group of methods were revised without arriving at a real conclusion (19). Since then, many sophisticated and sensitive biochemical procedures have been developed for the determination of even minimal levels of oxidative stress *in vivo* (19–25). More than 50 indices are in common use to assess oxidative stress, thus implementing more than 100 methods.

In my opinion the problem should be faced from two points of view in order to create a system for oxidative stress assessment that can serve as a biochemical control for medical ozone applications: (1) identification of appropriate biochemical indices and (2) interpretation of selected biochemical indices.

Identification of Appropriate Biochemical Indices

Critical substrates for ozone are lipids, proteins, carbohydrates and DNA; however, lipids and proteins are the main targets in the ozone reactions with blood plasma, rectal mucosa or biological fluids. The reaction with lipids occurs almost exclusively with the carbon-carbon double bonds present in unsaturated fatty acids, and the reaction with proteins is with the side-chain functionalities of several amino acid residues (26). These reactions produce different ozonation products (27–30) that have the task to relay the ozone effects to distant sites in the organism (31–33). However, when the amount of ozonation products overwhelms the antioxidant system, a toxic effect leading to tissue damage and diseases is established. Therefore, the expected therapeutic effects from ozone therapy depend on a careful measurement of the ozone dose to obtain an appropriate concentration of the ozonation products to have the desired effect with no toxicity (34,35). The mechanism of the products producing the therapeutic effect is not yet clear, but it is evident that a transient oxidative pulse acting as a cellular metabolic challenger is implicated. In other words, these oxidized products can act as signal transduction molecules to initiate a series of metabolic cascades that end with the called biological pulse due to ozone therapy.

The different ozonation products from lipids and proteins are Criegee ozonides, hydrogen peroxides, hydroxyhydroperoxides and aldehydes (27–30). To have an idea of the enzymatic antioxidant defense, one has to look inside the metabolic pathway of ozonation products. Criegee ozonides are catabolized by the enzymes glutathione S-transferases (GST) (36,37) using glutathione (GSH) as reduced cofactor, yielding aldehydes and oxidized glutathione (GSSG) (Figure 1). The glutathione S-transferases are a family of isoenzymes catalyzing the conjugation of glutathione with a large variety of xenobiotics as well as endogenous substrates (38–40). Aldehyde formation and formation of GSSG have been established by coupling of an aldehyde dehydrogenase assay or a glutathione reductase assay to the GST-catalyzed reaction of ozonides with GSH (41). Aldehydes are metabolized by the enzyme aldehyde

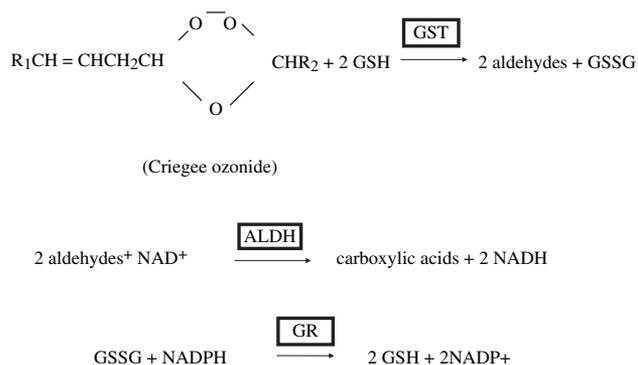


Figure 1. Metabolic pathway for Criegee ozonides and aldehydes.

dehydrogenase (ALDH) with oxidized nicotinamide adenine dinucleotide (NAD⁺) as cofactor (41) or the enzyme GST as occurs in the case of 4-hydroxynonenal using GSH as cofactor (42). GSH regeneration is achieved by glutathione reductase enzyme (GR) (Figure 1).

Hydroxyhydroperoxides (ROOH) are mainly metabolized through the enzyme glutathione peroxidase (GPx) with GSH as a reducing cofactor (Figure 2) and the regeneration of GSH is also obtained by the anaplerotic enzyme glutathione reductase. A stimulation of GPx activity during the application of systemic ozone therapy has been previously reported (43–46). However, organic hydroxyperoxides are also catalytically decomposed by the GST in the presence of a small amount of glutathione (47–49). Furthermore, the concentration of glutathione may regulate the catabolizing action of GPx or GST on hydroxyhydroperoxides. Hydrogen peroxide (H₂O₂) at a very low concentration is catalyzed by GPx but at high concentrations the catalase enzyme (Cat) is involved (50,51) (Figure 2).

The enzyme superoxide dismutase (SOD) catalyzes the dismutation of superoxide radical (O₂⁻) to hydrogen peroxide (52) (Figure 3). A simplified blood method to determine SOD has been developed in our laboratory (53). Although superoxide radical is not directly produced by ozone reaction with biomolecules, it must be taken into account because of its direct relation to form the hydroxyl radical (HO[•]) that is a more powerful oxidant. Ozone reaction forming hydroxyl radical has been proved in *in vitro* conditions (54), but until now it has not been detected in *in vivo* conditions.

On the other hand, the nonenzymatic antioxidant defense line must also be taken into account. In this context GSH has an antioxidant activity by itself because of the high reactivity of ozone with the cysteine residue in GSH molecule (55), and the antioxidant nutrients like α -tocopherol, ascorbic acid and β -carotene protect the cells against lipid peroxidation (56–59). Urate has also been reported to have an important antioxidant protective activity (60). All of these antioxidant substances, acting as radical scavengers, appear to be of vital importance in the protection of biological membranes against lipid peroxidation. A previous study showed that a significant increase in the content of blood GSH paralleled with a decrease in lipid peroxidation activity after ozone therapy treatment of rabbits (61).

The analysis allows us to assume that the changes in the distinct catabolic compounds of the ozonation products may serve as possible indices for oxidative stress estimation.

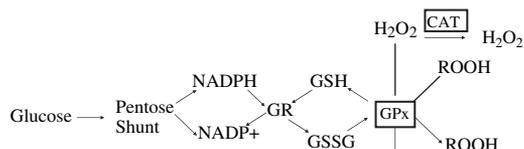


Figure 2. Metabolic pathway for hydroxyhydroperoxide and hydrogen peroxide.

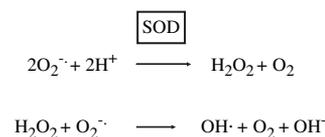


Figure 3. Equation for superoxide radical and hydrogen peroxide catabolism.

Nonetheless, the oxidation products from the different biomolecules should also be analyzed as possible biochemical indices. Thus, oxidant activity can be followed by means of the metabolites from lipid peroxidation reactions. The sequence of reactions during lipid peroxidation is shown in Figure 4. Lipid peroxidation is a process determined by the extent of peroxide-forming free radical mechanisms (62). Conjugated dienes (CD) arise as an early event of reactions of lipid peroxidation.

Several end products of lipid peroxidation such as aldehydes react readily with thiobarbituric acid (TBA) and are defined as thiobarbituric acid reactive substances (TBARS) serving as a measurement of tissue oxidative stress (63,64). Fluorescent chromolipids (FC) are inactive compounds formed by the reaction of aldehydes with free amino group in proteins and can easily be measured with a spectrofluorometer. In general, diene conjugation is a measure of early events of lipid peroxidation reactions. TBARS measures end products of lipid peroxidation and FC are products formed by reaction of end products of lipid peroxidation with the amino groups (65). The CD/FC ratio has been used as an effective measurement of peroxidation phase (2), but

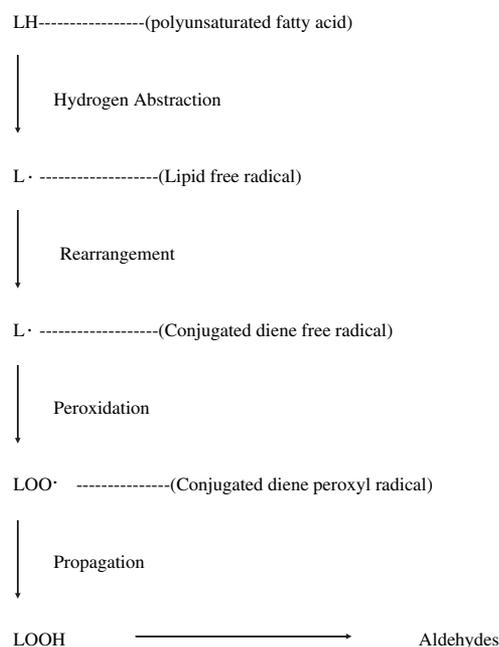


Figure 4. Figure showing the lipid peroxidation events.

its real effectiveness needs to be proven in different biological situations.

Proteins in plasma and interstitial fluids are also good substrates for ozone reaction during ozone therapy applications. In the case of free amino acids and the amino acids in proteins, cysteine, tryptophan, methionine, tyrosine, phenylalanine and histidine residues are particularly sensitive to oxidation by ozone; rate constants for reaction of ozone with these amino acids are 2–6 orders of magnitude larger than with leucine, valine and similar amino acids (26,55,66). Proteins undergoing oxidation generate a variety of products from ozone reaction with such amino acids; they are mainly Criegee ozonides and hydrogen peroxides with intermediate metabolites that are hydroxyhydroperoxides and aldehydes (66), similar to what was stated before for lipids, but ending the reactions in new products depending on the type of amino acid that undergo oxidation (67). Thus, oxidant activity in proteins can be evaluated by measurement of protein hydroperoxides, oxidized amino acids, protein carbonyl group content and protein-thiol group degradation (68–72).

There are several studies demonstrating that oxidative stress cannot be defined in universal terms because this is a biologically complex process that needs to be estimated from different viewpoints (73–75). Therefore, now it is universally accepted that assessment of oxidative stress status of an individual may require a combination of methods, each one representing a different type of oxidative stress because different pathologies may be associated with different types of oxidative stress and need to be evaluated by different indices (74).

Having in mind all of the before considerations and taking into account that the first line of defense is the most important against oxidative stress in humans, we have proposed eight possible relevant blood indices or biomarkers for assessing the real oxidative stress in a human (Table 1).

All of these biomarkers shown in Table 1 can be routinely measured using both blood red cells and plasma samples. The occurrence of compensatory homeostasis among different antioxidants in a given tissue can lead to erroneous conclusions regarding the physiological meaning of changes in the measurement of a particular antioxidant. Analyzing this battery of indices, one will have a better knowl-

edge on how ozone therapy should be applied and how it works in each patient.

Interpretation of Selected Biochemical Indices

Because of the complexity of the oxidative stress phenomenon, the analysis of the antioxidant and prooxidant indices cannot be simply done. In a simple manner, the increment in antioxidant parameters can be assumed as a positive or negative response to a given oxidant stimulus; on the contrary, the increment in prooxidant parameters can be assumed as a negative response. However, biological phenomena in a few cases are simple and in the majority of occasions are very complex with interrelationships between many particular biological events that give a resulting effect as final response. This is the key point in the way of interpretation of the different indices or parameters to assess the oxidative status. Many authors have patented different methods for the analysis of the different indices measured for assessing the oxidative stress. Some of them based their analysis by comparison to reference values from apparently normal donors, others by making diagnostic graphic plots of a huge number of assayed parameters, and others by using cells isolated in a noninvasive assay (76–79). All of these diagnostic systems have their advantages and disadvantages, depending on the feasibility of the user. These systems apparently give an idea if an oxidative stress exists, but none is really able to predict the grade or level of redox status. For ozone therapy applications it is very important to know the patient's level of oxidative stress.

An example on how interpretation from antioxidant and prooxidant parameters is done at present is shown in Figure 5. These values were obtained from a cardiac patient who suffered myocardial infarction 6 months previously. He was treated with ozone major autohemotherapy with a protocol comprising volume of blood 200 mL, ozone concentration 50 µg/mL/mL of blood, at a frequency of 2 or 3 days a week.

Table 1. Biochemical indices for a proposed system for measuring the oxidative stress status in humans from a blood sample

Antioxidants	Prooxidants
Reduced glutathione (GSH)	
Glutathione peroxidase (GPx)	Conjugated dienes (CD)
Glutathione S-transferase (GST)	Total hydroperoxides (THP)
Superoxide dismutase (SOD)	Thiobarbituric acid reactive substances (TBARS)
Catalase (CAT)	

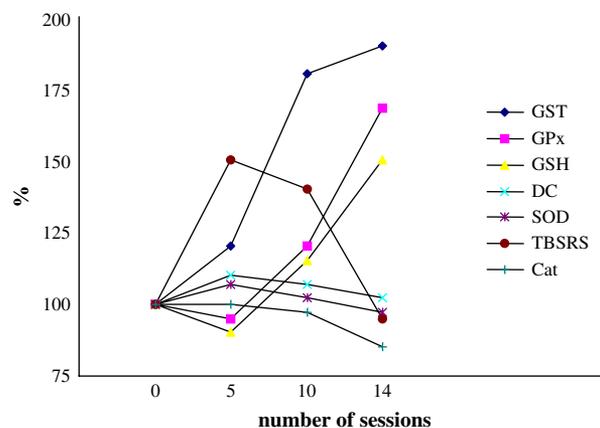


Figure 5. Response of antioxidant and prooxidant parameters during ozone major autohemotherapy application in a cardiopathic patient.

The response was plotted in terms of percent vs. number of ozone therapy sessions. Before starting the therapy, blood sample was drawn and prooxidant and antioxidant values were analyzed, corresponding to 100% of each parameter.

After five sessions of treatment, blood analysis was repeated and compared with the first one in order to determine the individual specific changes after ozone treatment. There was a slight increase in the prooxidant state, given by a high level of TBARS and a small increase in DC, along with a small increase in the antioxidant enzyme GST but no significant changes in GPx, Cat and SOD with a diminished level in GSH. Evidently, there were uncertain changes. After 10 treatment sessions, a significant increase in the antioxidant parameters GST, GPx and GSH with a decay in SOD and Cat was observed; the prooxidant indices DC and TBARS were slightly decreased in comparison with the prior analysis. Results were similar at the end of 14 sessions but with a high increase in GPx and GSH, and a high decrease in CAT and TBARS indices. Although it looks like an induction in antioxidant activities occurred at the end of the treatment, there was no clear evidence or interpretation of this phenomenon because many biological interrelations were not taken into account. This is the main reason why I am stating that ozone therapy needs a control system including all possible biological aspects related to oxidative stress. Therefore, our laboratory developed a computer algorithm taking into account all the biological relationships between antioxidant and prooxidant activities, which gives a total measurement of both activities with a redox index and grade of oxidative stress.

Proposal of a Novel System for Oxidative Stress Diagnosis

There is growing interest by scientists, ozone therapy and clinical medicine specialists for a reliable and cost-effective system in which the interpretation of the included parameters is effective in realistically estimating changes and level of oxidative stress in humans.

A system based on the technique of neuronal artificial net has been developed in our laboratory for the interpretation of the proposed indices. This new technique is based on the integration of different but complementary biochemical parameters that cover the main aspect of the biological oxidative stress. A computer program has been constructed which yields four new relevant indices for final oxidative stress diagnosis. These indices are shown in Table 2.

Table 2. Resulting indices from oxidative stress diagnostic computer program

Total antioxidant activity
Total prooxidant activity
Redox index
Grade

Table 3. Grades of oxidative stress and their clinical meaning

Grade	Meaning
0	No oxidative stress
1	Light oxidative stress
2	Moderate oxidative stress
3	Severe oxidative stress
4	Very severe oxidative stress

Total antioxidant activity index characterizes the level of the antioxidant capacity of the whole organism, taking into account the enzyme and nonenzyme antioxidant defenses. Total prooxidant activity index indicates the level of oxidative activity to which the cells are submitted. Redox index represents the numerical value of the resulting vector from both activities. Grade index displays the level of oxidative stress that the organism is undergoing and is composed of five grades indicating five clinical oxidative statuses (Table 3).

Although HPT biomarker was not measured in the case of Figure 5, we made an arrangement in the computer program to evaluate the results from the biomarkers analyzed in the blood of that patient before, during and after ozone therapy applications. As can be seen in Table 4, the level of oxidative stress status in each point was possible to be assessed without any doubts or misinterpretations.

A slight increase in the oxidative stress level was achieved after five sessions of treatment; however, a significant increase in the total antioxidant activity produced a remarkable increase in the redox index after ten sessions, with a minor level of oxidative stress grade. It is evident that an improvement in the oxidative stress status at the end of 14 sessions occurred because the patient had one grade less than before treatment. Clinical status was well correlated with the redox status given by the program.

Many of the oxidized damage components produced throughout the body are transported to the blood plasma, urine or breath; thus, analyzing the oxidative level of plasma is a measure of possible oxidant attack at which cells have to face. By contrast, analysis of the antioxidant level inside the red cell is a measure of the defense capacity against the oxidative action outside the cell. These concepts

Table 4. Oxidative stress diagnosis using the computer program for values obtained with the patient in Figure 5

Ozone therapy session number	Total antioxidant activity (units)	Total prooxidant activity (units)	Redox index	Grade of oxidative stress
0	128.38	167.96	0.764	2
5	123.45	178.69	0.691	3
10	138.92	175.21	0.790	2
14	144.28	169.70	0.850	1

are undertaken by the program along with the interrelationship between each group of indices and the crossover between them.

This oxidative stress diagnostic system has the advantage that does not only give a measure if a person is suffering from oxidative stress, it additionally shows the level of oxidative status that is a very relevant point because it allows the physicians to control the ozone doses and safety of the therapy.

In conclusion, oxidative stress phenomenon is a very important biological process that all ozone therapists should have in mind when using systemic ozone therapy: first, because this therapy works by mean of a transient and controlled exogenous oxidative stress produced from a myriad of ROS and second, because the transient oxidative stress needs to be controlled by the ozone dosage in order not to cause toxic or only placebo effects. And third, and most important, the redox status of the patient should be assessed before, during and after systemic ozone therapy applications.

A reliable system that allows knowing the patient's redox status and estimation of the oxidative stress level has been developed in our laboratory, with implications on the dosage and therapeutic effectiveness of ozone therapy.

Ozone therapists should have in mind that safe dosage in ozone therapy is not always the therapeutic dosage; hence, biochemical findings in relation to the patient's ailment, therapy objective and pro-oxidant/antioxidant balance should be analyzed as an entire system for safety and therapeutic actions. Interpretation of these parameters is the most important part of any system evaluating the oxidative level in a patient. Decay in the oxidant state and/or the increase in the antioxidant state is not only a homeostatic response of the cells, but also represents the safety and effectiveness of applied ozone dose. For that reason, a biochemical control system will be very useful in certification of the optimal use of ozone therapy applications, although any measurement, given an idea of the oxidative stress status, can be used. The real proposal is that when using systemic ozone therapy, one control system of the oxidative stress must always be applied.

References

- Bocci B. Does ozone therapy normalize the cellular redox balance? *Med Hypoth* 1996;46:150–154.
- Kontorschikova CN. Biochemical safety control in ozone therapy. *Proceedings of the 12th World Congress of the International Ozone Association (Vol. 3)*. Lille, France; 1995. pp. 231–234.
- Bocci V. The antioxidant system and the defence system against ozone. In: Bocci V, ed. *Oxygen-Ozone Therapy. A Critical Evaluation*. Dordrecht: Kluwer Academic Publishers; 2002. pp. 79–107.
- Sies H. Oxidative stress: introductory remarks. In: Sies H, ed. *Oxidative Stress*. London: Academic Press; 1985. pp. 18–28.
- Sies H. Oxidative stress: from basic research to clinical application. *Am J Med* 1991;91:3C/31S–3C/38S.
- Farber JL, Kyle ME, Coleman JB. Mechanism of cell injury by activated oxygen species. *Lab Invest* 1990;62:670–679.
- Janssen YMW, Van Houten B, Borm PJA, Mossman BT. Cell and tissue response to oxidative damage. *Lab Invest* 1993;69:261–274.
- Wiseman H, Halliwell B. Damage to DNA by reactive oxygen and nitrogen species: role in inflammatory disease and progression to cancer. *Biochem J* 1996;313:17–29.
- Poli G. Liver damage due to free radicals. *Br Med Bull* 1993;49:604–620.
- Powis G, Briehl M, Oblong J. Redox signaling and the control of cell growth and death. *Pharmacol Ther* 1995;65:149–173.
- Lander HM. An essential role for free radicals and derived species in signal transduction. *FASEB J* 1997;11:118–124.
- Nakamura H, Nakamura K, Yodoi J. Redox regulation of cellular activation. *Annu Rev Immunol* 1997;15:351–369.
- Sen CK. Redox signaling and the emerging therapeutic potential of thiol antioxidants. *Biochem Pharmacol* 1998;55:1747–1758.
- Ishii T, Itho K, Ruiz E, Bannai S, Yamamoto M, Mann GE. 4-Hydroxynonenol induces expression of antioxidant stress proteins via the transcription factor Nrf2 in cultured murine macrophages and vascular smooth muscle cells: lack of heme oxygenase-1, peroxiredoxin I and A170 induction in nrf2-knockout mice. *J Physiol* 2001;531P:9.
- Conner E, Miles AM, Aiko A, Grisham MB. Reactive oxygen metabolites. Therapeutic implications of their role in inflammatory diseases. *Cell Immunother* 1995;3:438–449.
- Halliwell B, Gutteridge LMC, eds. *Free Radicals in Biology and Medicine*. 2nd ed. Oxford: Clarendon Press; 1989.
- Favier A. Oxidative stress: value of its demonstration in medical biology and problems posed by the choice of markers. *Ann Biol Clin* 1997;55:9–16.
- Pryor WA, Godber SS. Oxidative stress status: an introduction. *Free Radic Biol Med* 1991;10:173.
- Pryor AA, Godber SS. Noninvasive measures of oxidative stress status in humans. *Free Radic Biol Med* 1991;10:177–184.
- Packer L. Oxygen radicals in biological systems. *Methods Enzymol* 1994;233. (special issue).
- Rice-Evans C, Miller BJ. Total antioxidant status in plasma and body fluids. *Methods Enzymol* 1994;234:279–293.
- Nourooz-Zadeh J, Tajaddini-Sarmadi J, Wolf SP. Measurement of plasma hydroperoxide concentrations by the ferrous-oxidation xylenol orange (FOX) assay in conjunction with triphenylphosphine. *Anal Biochem* 1994;220:403–409.
- Alberti A, Bolognini L, Carratelli M, Della Bona MA, Macciantelli D. Assessing oxidative stress with the d-ROMs test. Some mechanistic considerations. *Proceedings of the SFRR Europe Summer Meeting 26–28 June*. Abana Terme, Italy; 1997. pp. 82–83.
- Ahotupa A, Marniemi J, Lehtimäki T, Talvinen K, Raitakari OT, Vasankari T, et al. Baseline diene conjugation in LDL lipids as a direct measure of in vivo LDL oxidation. *Clin Biochem* 1998;31:257–261.
- Levine RI, Wehr N, Williams JA, Stadman ER, Schacter F. Determination of carbonyl groups in oxidized proteins. *Methods Mol Biol* 2000;99:15–24.
- Uppu RM, Pryor WA. The reactions of ozone with proteins and unsaturated fatty acids in reverse micelles. *Chem Res Toxicol* 1994;7:47–55.
- Pryor WA, Wu M. Ozonation of methyl oleate in hexane, in a thin film, in SDS micelles, and in distearoylphosphatidylcholine liposomes: yields and properties of the Criegee ozonide. *Chem Res Toxicol* 1992;5:505–511.
- Pryor WA, Uppu RM. A kinetic model for the competitive reactions of ozone with amino acid residues in proteins in reverse micelles. *J Biol Chem* 1993;268:3120–3126.

29. Squadrito G, Uppu RM, Cueto R, Pryor WA. Production of the Criegee ozonide during the ozonation of 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine liposomes. *Lipids* 1992;27:955–958.
30. Uppu RM, Cueto R, Squadrito GL, Pryor WA. What does ozone react with at the air/lung interface? Model studies using human red blood cell membranes. *Arch Biochem Biophys* 1995;319:257–266.
31. Pryor WA, Squadrito GL, Friedman M. The cascade mechanism to explain ozone toxicity: the role of lipid ozonation products. *Free Radic Biol Med* 1995;19:935–941.
32. Pryor WA, Squadrito GL, Friedman M. A new mechanism for the toxicity of ozone. *Toxicol Lett* 1995;82:287–293.
33. Wright DT, Friedman M, Pryor WA, Squadrito GL, Salgo MG, Adler KB. Lipid ozonation products activate phospholipases in guinea pig and human airway epithelial cells in vitro. *Am J Resp Crit Care Med* 1994;49:A320.
34. Bocci V. A reasonable approach for the treatment of HIV infection in the early phase with ozonotherapy (autohemotherapy). How inflammatory cytokines may have therapeutic role. *Mediators Inflamm* 1994;3:315–321.
35. Bocci V. Ozone: a mixed blessing. *Res Complementary Med* 1996;3:25–33.
36. Rietjens IM, Lemmink HH, Alink GM, van Bladeren PJ. The role of glutathione and glutathione S-transferases in fatty acid ozonide detoxification. *Chem Biol Interact* 1987;62:3–14.
37. Hempenius RA, de Vries J, Rietjens IMCM. Molecular orbital study on the glutathione-dependent detoxification of ozonides. *Eur J Pharmacol Environ Toxicol Pharmacol Sect* 1992;228:207–211.
38. Commandeur JNM, Stijntjes GJ, Vermeulen NPE. Enzymes and transport systems involved in the formation and disposition of glutathione S-conjugates. *Pharmacol Rev* 1995;47:271–330.
39. van Bladeren PJ, van Ommen B. The inhibition of glutathione S-transferases: mechanisms, toxic consequences and therapeutic benefits. *Pharmacol Ther* 1991;51:35–46.
40. Jakoby WB. The glutathione S-transferase: a group of multifunctional detoxification proteins. *Adv Enzymol* 1978;46:383–414.
41. Vos RME, Rietjens IMCM, Stevens LH, Bladeren PJV. Methyl linoleate ozonide as a substrate for rat glutathione S-transferases: reaction pathway and isoenzyme selectivity. *Chem Biol Interact* 1989;69:269–278.
42. Xie C, Lovell MA, Markesbery WR. Glutathione transferase protects neuronal cultures against four hydroxynonenal toxicity. *Free Radic Biol Med* 1998;25:979–988.
43. Hernández F, Menéndez S, Gómez M, Eng L. Efecto de la ozonoterapia intravascular sobre el sistema de la glutatión peroxidasa. *Rev CENIC Ciencias Biol* 1989;20:37–40.
44. Hernández F, Menéndez S, Wong R. Decrease of blood cholesterol and stimulation of antioxidative response in cardiopathy patients treated with endovenous ozone therapy. *Free Radic Biol Med* 1995;19:115–119.
45. Hernández F, Calunga JL, Turrent J, Menéndez S, Montenegro A. Ozone therapy effects on biomarkers and lung function in asthma. *Arch Med Res* 2005;36:549–554.
46. Larini A, Bianchi L, Bocci V. The ozone tolerance. I. Enhancement of antioxidant enzymes is ozone dose-dependent in Jurkat-cells. *Free Radic Res* 2003;37:1163–1168.
47. Beckett GJ, Hayes JD. Glutathione S-transferases: biomedical applications. *Adv Clin Chem* 1993;30:281–380.
48. Aniya Y, Anders MW. Regulation of rat liver microsomal glutathione S-transferase activity by thiol/disulfide exchange. *Arch Biochem Biophys* 1989;270:330–334.
49. Aniya Y, Daido A. Organic hydroperoxide-induced activation of liver microsomal glutathione S-transferase of rats in vitro. *Jpn J Pharmacol* 1993;62:9–14.
50. Scott MD, Lubin BH, Zuo L, Kuypers FA. Erythrocyte defense against hydrogen peroxide preeminent importance of catalase. *J Lab Clin Med* 1991;118:7–16.
51. Eaton JW. Catalases and peroxidases and glutathione and hydrogen peroxide: mysteries of the bestiary. *J Lab Clin Med* 1991;118:3–4.
52. Fridovich I. Superoxide dismutase. *Annu Rev Biochem* 1975;44:147–159.
53. Hernández F, Prats M, González M. Un método simplificado para la determinación de superóxido dismutasa. *Rev CENIC Ciencias Biol* 1994;25:27–29.
54. Pryor WA. Mechanism of radical formation from reactions of ozone with target molecules in the lung. *Free Radic Biol Med* 1994;17:451–465.
55. Pryor WA. How far does ozone penetrate into the pulmonary air/tissue boundary before it reacts? *Free Radic Biol Med* 1992;12:83–88.
56. Watanowicz M, Panczenko-Kresowska B, Ziemblanski S, Kowalska M, Okolska G. The effect of α -tocopherol and ascorbic acid on the serum lipid peroxide level in elderly people. *Ann Nutr Metab* 1984;28:186–191.
57. Vatassery GT, Smith WE, Quach HT. Ascorbic acid, glutathione and synthetic antioxidant prevent the oxidation of vitamin E in platelets. *Lipids* 1989;24:1043–1047.
58. Pryor WA. The antioxidant nutrients and disease prevention—what do we know and what do we need to find out? *Am J Clin Nutr* 1991;53:391S–393S.
59. Kaneko T, Kaji K, Matsuo M. Protective effect of lipophilic derivatives of ascorbic acid on lipid peroxide-induced endothelial injury. *Arch Biochem Biophys* 1993;304:176–180.
60. Frei B, Stocker R, Ames BN. Antioxidant defenses and lipid peroxidation in human blood plasma. *Proc Natl Acad Sci USA* 1988;85:9748–9752.
61. Hernández F, Menéndez S, Alvarez I. Blood and tissue biochemical study of normo- and hypercholesterolemic rabbits treated with ozone. *Proceedings of the 12th World Congress of the International Ozone Association*. Lille, France; 1995. pp. 251–255.
62. Dormandy T. Free-radical oxidation and antioxidants. *Lancet* 1978;1:647–650.
63. Gutteridge JMC, Halliwell B. The measurement and mechanism of lipid peroxidation in biological systems. *TIBS* 1990;15:129–135.
64. Valenzuela A. The biological significance of malondialdehyde determination in the assessment of tissue oxidative stress. *Life Sci* 1991;48:475–477.
65. Vasankari T, Kujala U, Heinonen O, Kapanen J, Ahotupa M. Measurement of serum lipid peroxidation during exercise using three different methods: diene conjugation, thiobarbituric acid reactive material and fluorescent chromolipids. *Clin Chim Acta* 1995;234:63–69.
66. Berlett BS, Levine RL, Stadtman ER. Comparison of the effects of ozone on the modification of amino acid residues in glutamine synthetase and bovine serum albumin. *J Biol Chem* 1996;271:4177–4182.
67. Stadtman ER. Protein oxidation and aging. *Science* 1992;257:1220–1224.
68. Wolf SP. Ferrous ion oxidation in the presence of the ferric ion indicator xylenol orange for the measurement of hydroperoxides; the FOX assay. *Methods Enzymol* 1994;233C:182–189.
69. Nourooz-Zadesh J, Tajaddini-Sarmadi J, Wolf SP. Measurement of plasma hydroperoxide concentrations by the ferrous-oxidation xylenol orange (FOX) assay in conjunctions with triphenylphosphine. *Anal Biochem* 1994;220:403–409.
70. Stadtman ER, Oliver CN. Metal catalyzed oxidation of proteins. Physiological consequences. *J Biol Chem* 1991;256:2005–2008.
71. Stadtman ER, Levine RL. Protein oxidation. *Ann NY Acad Sci* 2000;899:191–208.
72. Hu ML. Measurement of protein thiol groups and glutathione in plasma. *Methods Enzymol* 1994;233:380–385.
73. Arab K, Steghens JP. Plasma lipid hydroperoxides measurement by an automated xylenol orange method. *Anal Biochem* 2004;325:158–163.

74. Dotan Y, Lichtenberg D, Pinchuk I. Lipid peroxidation cannot be used as a universal criterion of oxidative stress. *Prog Lip Res* 2004;43:200–227.
75. Arguelles S, García S, Maldonado M, Machado A, Ayala A. Do the serum oxidative stress biomarkers provide a reasonable index of the general oxidative stress status? *Biochim Biophys Acta* 2004;1674:251–259.
76. Heinecke JW. Method for the determination of oxidative stress. USA Patent 2000; 6096556.
77. Kinkade JM Jr, Brown WV, Pohl J, Shapira R, Jensen PE, Le NA. Biomarkers for oxidative stress. PCT Patent WO 2000; 2000–028072.
78. Lamb RG. Test for oxidative stress using cell suspensions. PCT Patent WO 2000; 2000–000809.
79. Ochi H, Cutler R. Use of oxidative diagnostic plot as a health indicator for assessing oxidative stress and its control in humans. PCT Patent WO 1999; 1999–063341.